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BROWN ROOT ROT DISEASE IN AMERICAN SAMOA

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ABSTRACT

Phellinus noxius causes root and lower stem rot of woody plants throughout the South Pacific region. Its hosts include rubber, mahogany, cacao, and many timber, fruit and landscape trees. Though endemic to the tropics, we found no reports describing brown root rot disease in native forests, exclusively. Incidence, distribution and host range of *P. noxius* were measured in primary and secondary rainforests on Tutuila Island, American Samoa. *P. noxius* was recorded in 19 of 20 strip transects and 1.2 ha established plots and in all vegetation types, infecting 37 tree species in 30 genera and 22 families. Species most affected were *Myristica fatua*, *Dysoxylum samoense* and *Hibiscus tiliaceus*^{3/4}25, 16 and 10 percent, respectively. Of 62 infection centers, 33 contained the same tree species and 13 were dominated by a single species. The fewest infections were recorded at primary montane and ridge top sites, the most in secondary valleys. Disease incidence was influenced more by human disturbance than by vegetation type, topography, stem diameter, stem density, or soil type. Regenerating secondary valley sites appeared to lack the species richness of mature sites, had the highest disease incidence, and the largest and highest number of infection centers. This agrees with other host/pathogen associations, such as Douglas-fir/*P. weirii* and hardwood/*P. noxius* plantations, where disease incidence and spread was higher in species poor than in species rich stands.

INTRODUCTION

Brown root rot disease, caused by *Phellinus noxius* (Corner) Cunningham, has been reported throughout the Pacific and Southeast Asia as a cause of tree decline and mortality (McKenzie 1996). The name brown root rot refers to a brown to black mycelial crust formed by the fungus on the surface of infected roots and stem bases (Chang & Yang 1998). However, *P. noxius* is generally considered a white rot fungus because

of its ability to degrade lignin, a basic component of wood (Adaskaveg & Ogawa 1990, Chang & Yang 1998).

Most of the literature on *P. noxius* describes root and crown rot of plantation crops such as hoop pine (*Araucaria cunninghamii* Ait. ex D. Don; Bolland 1984), cacao (*Theobroma cacao* L.; Thrower 1965), rubber (*Hevea brasiliensis* (Willd. ex Adr. de Juss) Muell. Arg.; Nandris et al. 1987), mahogany (*Swietenia macrophylla* King; Singh et al. 1980) and *Cordia alliodora* (Ruiz. and Pav.) Oken (Neil 1986). Exotic trees introduced in reforestation projects and infected by *P. noxius* include *C. alliodora* (Neil 1986), *S. macrophylla* and *Gmelina arborea* Roxb. (Ivory 1990). Woody perennials in fruit orchards and landscape plantings have also been documented as susceptible to brown root rot (Hodges & Tenorio 1984, Chang 1995, Ann et al. 1999) and Chang and Yang (1998) list 60 fruit, forest and ornamental plant species infected by the fungus in Taiwan. We found no published reports limited to *P. noxius* damage in native tropical forests.

Presently in American Samoa there is little concern over occasional fruit or landscape trees killed by brown root rot disease. Many farmers have lost breadfruit trees (*Artocarpus altilis* (S. Parkinson) Fosb.) to the disease, first reported in American Samoa as charcoal crown rot, caused by *Corticium* sp. (Trujillo 1971), but later recognized as *P. noxius* by Hodges and Tenorio (1984). Due to limited land and a rapidly increasing population, however, farmers are moving up valleys and ridges into native forests (Volk et al. 1992). *Phellinus noxius* is present in the forests of American Samoa (F. Brooks pers. obs.) and recently cleared planting sites may be infested with the fungus, causing death of agroforest species, including breadfruit and citrus. *Phellinus noxius* spreads by root contact and may persist in roots and stumps of infected plants for more than 10 years after death of the host (Chang 1996). The purpose of this study was to determine the incidence, distribution and host range of *P. noxius* in

primary and secondary forests on Tutuila, the main island of American Samoa.

MATERIALS AND METHODS

STUDY SITES

All sites were on Tutuila Island, located at 14°18' S latitude and 170°41' W longitude. Average annual temperature is 26.4°C and annual rainfall ranges from 2500 mm on the Tafuna Plain to over 6000 mm on Matafao Peak. Tutuila's eroded volcanic ridges and valleys cover 137 km², 65 percent of which consist of steep (>30%) heavily forested slopes (Wingert 1981). Slopes in most rainforests are from 40-70 percent with areas exceeding 100 percent (Webb & Fa'aumu 1999). Most of the Territory's estimated 65,000 inhabitants live on Tutuila.

Twenty survey sites were selected that were both accessible and large enough for transects. They included five primary (late successional) vegetation types: montane forest, ridge, slope, and valley lowland forest, littoral forest, and Tafuna lowland forest (Whistler 1994). Three examples of secondary (disturbed or early successional) vegetation were also surveyed. The total area sampled was 8.8 ha, of which 1.2 ha was secondary forest and 7.6 ha primary forest.

The littoral forest, on or just inland from the coast, is characterized by *Barringtonia asiatica* and other salt-tolerant plant species dispersed by the sea or by ocean birds (Cole et al. 1988). Lowland forest, also called tropical rainforest or high forest, covers most of the island and is divided by Whistler (1994) into coastal, valley and ridge vegetation types found below 350 m. Differences in soil, topography, elevation and natural and man-made disturbances affect these vegetation types (Whistler 1994, Webb et al. 1999). Lowland forest tree species include *Syzygium inophylloides*, *Calophyllum neo-ebudicum*, *Canarium vitiense*, *Diospyros samoensis*, *Canarium harveyi* and *Myristica fatua*. Montane forest occurs at elevations above 350 m and

is periodically altered by human or natural events, such as hurricanes and landslides. Predominant montane species include *Dysoxylum huntii*, *Syzygium samoense* and *Crossostylis biflora*.

The survey also included the Tafuna Plain lowland forest, a remnant of tropical forest that once covered most of the large, relatively flat Tafuna Plain. Dominant tree species in the Tafuna forest are *Planchonella samoensis*, *Pometia pinnata* and *Dysoxylum samoense* (Webb et al. 1999). Though reduced to approximately 8 ha and surrounded by human habitation, almost 20 percent of the 425 indigenous angiosperms in American Samoa occur in this species rich community (Whistler 1993). Further, since large areas of the Plain are still agroforest, the incidence of *P. noxius* in this remaining stand was of interest. The Tafuna lowland forest is also a target of renewed conservation efforts based on the work of Volk et al. (1992), Whistler (1993) and others.

In order to determine if the incidence and distribution of *P. noxius* varied in disturbed areas, secondary forest sites (Whistler 1994) were included in the survey. Key tree species indicative of forest disturbance include *Rhus taitensis*, *Hibiscus tiliaceus*, *Macaranga harveyana* and *M. stipulosa*. (Webb et al. 1999).

SOILS

Descriptions of soil types were based on work by the USDA Soil Conservation Service (Nakamura 1984). Soils at most sites (12/18) are derived from Fagasa family-Lithic Hapludolls-Rock outcrop (Table 1). Approximately 55 percent of this soil unit, the Fagasa family, is found on ridges and slopes from 70-130 percent. They are formed from igneous bedrock, are well drained and fairly deep. Lithic Hapludolls are cobbly and clayey, shallow and well drained and form 20 percent of this soil type. Very steep to vertical outcroppings of igneous rock make up another 15 percent of this unit, followed by small areas of talus, landslides and other soils. A very stony silty clay loam was present at four of the survey sites, on 30-60 per-

cent slopes and below 150 m. Permeability, as well as runoff, is rapid and the danger of erosion is severe. The soil unit at two disturbed sites in Malaeimi Valley was Leafu silty clay. This deep, alluvial, poorly drained soil is generally found on valley floors and near streams. Water tables tend to be high, 75-150 cm, leading to wet soils and occasional flooding. In the Tafuna Plain lowland forest the soil unit covering the lava beds is Tafuna extremely stony muck. It is deep, well drained, and broken by extensive outcroppings of lava stones. Soils at the Pago Pago Airport, where the Tafuna Plain meets the sea, are Troporthents. They have been disturbed by cutting, grading and filling and consist of sand, gravel, cobbles, and fine textured soil. Water erosion is minimal due to moderate soil permeability and slow to medium runoff.

SURVEY METHODS

Incidence and distribution of *Phellinus noxius* was determined by strip transects (Greenwood 1996) and fixed plots (Webb & Fa'aumu 1999). At least two strip transects were surveyed in each primary vegetation type by pacing off 245 meters with a measuring wheel (Measure Meter, model MM-50, by Rolatape, Spokane, Washington, USA). Workers examined every tree within five meters on either side of the wheel, producing a 2,450 m² (0.25 ha) transect. Information collected for all surveys included tree species with signs of *P. noxius* infection, tree location within transects, infection centers, and tree diameter at breast height (dbh). Infection centers were defined as two or more infected trees, each within 3 m of the nearest infected tree.

Four permanent American Samoa Government Department of Marine and Wildlife Resources (DMWR) research plots were also surveyed for *P. noxius*. These plots, located at Amalau, Vatia, Maloata and Alava, were established in 1998 by Webb and Fa'aumu (DMWR unpublished). Plots measured 100 m x 120 m (1.2 ha) and were sited to include ridge top, slope and valley lowland forest habitats (Table 1). All sites except Maloata included a stream bottom and the Amalau site

was located below ridge line. Trees in these plots with a dbh ³10 cm have been identified, measured and tagged. This provided information on stand density, species richness and distribution, and diameter class for comparison with data collected in our surveys. Spatial distribution of *P. noxius* in DMWR plots was analyzed using Spatial Point Pattern Analysis (SPPA ver. 1.1.1; Haase 2000).

PATHOGEN

Trees were considered infected by *P. noxius* if two or more of the following signs were present. On most trees a thick mycelial crust enveloped infected roots and the base of the stem up to 2 m (Bolland 1984, Hodges & Tenorio 1984, Ivory 1990). The white, expanding margin of the crust was associated with underlying sapwood discoloration (Thrower 1965, Singh et al. 1980, Hodges & Tenorio 1984, Nandris et al. 1987). Fine mats of mycelium were present between infected bark and sapwood and underlying colonized heartwood eventually became white, spongy, dry, and honeycombed with reddish-brown or dark lines (Singh et al. 1980, Nandris et al. 1987, Ivory 1990, Chang 1995, Nicole et al. 1995). Fruiting bodies (sporocarps) were occasionally present on standing and fallen trees. They were either shelf-like (dimidiate), growing flat along the undersides of fallen trees (resupinate), or a combination of both (effused-reflexed). The sterile upper surface of sporocarps was medium brown to black, rough and irregularly zoned. The fertile pore surface was usually gray-brown to umbrinous (Corner 1932, Fidalgo 1968, Pegler & Waterston 1968).

Phellinus lamaensis (Murrill) Heim is also present in the Samoas (McKenzie 1996). This fungus may occasionally be distinguished from *P. noxius* in the field by the ochre-colored expanding margin of the sporocarp; the margin of *P. noxius* sporocarps is usually creamy white (Corner 1932). Microscopically, *P. lamaensis* has conical, reddish-brown hymenial setae projecting conspicuously into the pores; hymenial setae are absent in *P. noxius* (Corner 1932, Fidalgo 1968, Pegler & Waterston 1968).

RESULTS

DISTRIBUTION AND FREQUENCY

Brown root rot disease was observed at 19 of the 20 sites surveyed on Tutuila Island (Table 1). It was not detected in a strip transect next to the Pago Pago Airport runway, a disturbed area dominated by *H. tiliaceus* and *Leucaena leucocephala* (Lam.) de Wit. *Phellinus noxius* was most prevalent in the three secondary (disturbed) valley sites, comprising 41 percent of all infections. The Malaeimi Valley soil (Table 1) is poorly drained silty soil with areas that remain wet for much of the year. Few infected trees were observed in the wettest areas of the Malaeimi II transect (Figure 1a, 120 m to 245 m) but *P. noxius* was heavily distributed along a dry streambed of loose, rocky soil in the Malaeimi I transect (Figure 1b, 160 m to 245 m). The Maloata Valley transect was perpendicular to and bisected by a perennial stream. Though *P. noxius* was recorded near the stream, most of the infected trees were located on steep slopes away from water (Figure 1c).

The Tafuna Plain lowland forest transects had 10 percent of the infected trees in strip surveys (Figure 1d), followed by littoral (9%), primary valley (8%), slope (7%), ridge top (4%) and montane (3%) sites. The disturbed ridge and airport were single transect sites.

HOST RANGE

Twenty-five of 324 infected trees from all sites could not be identified, mainly due to their advanced state of decay. The 299 identified host trees included 37 species in 30 genera and 22 families (Table 2). Of the 299 infected, identified trees at all sites, *M. fatua* was the most commonly affected (25%), followed by *D. samoense* (16%) and *H. tiliaceus*, (10%) (Table 2). We did not identify all trees in the 16 strip transects, but in the four DMWR plots *M. fatua* was the most abundant (31%), followed by *H. tiliaceus* (14%) (DMWR unpublished.). *D. samoense*, common in young secondary forests (Whistler 1994), com-

prised only 0.4% of tree species in the four primary forest (DMWR) plots (Webb & Fa'aumu 1999) but accounted for over 30 percent (35 of 150 trees) of the infected trees in two secondary valley sites.

INFECTION CENTERS

Sixty-one percent of all trees with brown root rot disease were in clusters of 2-10 infected trees. The distribution of infected trees was significantly clumped in all DMWR plots except Alava, according to Ripley's K-function based on a Monte-Carlo simulation with 1,000 iterations (Ripley 1981, Haase 1995). Of the 62 infection centers in both permanent and strip transect plots, 53 percent contained trees of the same species and one species was in the majority in 19 percent of the centers. For example, one center of eight infected trees at a disturbed valley site (Figure 1b) contained only *H. tiliaceus* and the four infection centers in the Tafuna Plain transects each consisted of a single tree species. The highest number of infection centers occurred in the disturbed valley sites: Malaeimi I, Maloata and Malaeimi II, with 10, 10, and 5 centers, respectively. Malaeimi I and Maloata had infection centers with the greatest number of trees (Figure 1b, 1c). The center covering the largest area, 100 m², was also in Malaeimi I.

DIAMETER CLASS

The total number of trees with a dbh ³10 cm was determined for the DMWR plots Alava, Amalau and Vatia (Webb & Fa'aumu 1999). Trees with a dbh >30 cm comprised 19 percent of all trees but 44 percent of infected trees (Table 3). Trees in strip transects were not counted and only the dbh of infected trees was measured: 26 percent of infected trees were in diameter classes >30 cm and 24 percent had a dbh <10 cm.

PATHOGEN

Characteristics of *P. noxius* sporocarps were within limits described by Corner (1932) and Fidalgo (1968). A mycelial crust approximately 1.0 cm thick was present on the upper roots, lower stem, or both, of most infected trees and was gen-

erally between 0.5-1.0 m in height. At the Malaeimi II site, however, crusts measured 3.0 m, 4.3 m and 4.85 m high on *M. fatua*, *C. odorata* and *D. samoense*, respectively. Microscopic examination revealed the reddish-brown lines in decayed wood to be both free and agglutinated fungal hyphae (Corner 1932).

DISCUSSION

Brown root rot disease is widely distributed in the tropical forests of Tutuila Island, American Samoa. It was found at all elevations and in all vegetation types and survey sites except one young, low diversity stand of *H. tiliaceus* and *L. leucocephala* next to the Pago Pago Airport runway. The disease was discovered, however, in a less disturbed continuation of the same forest outside the airport boundaries. Construction activities, including heavy grading and filling, may have disturbed or destroyed colonies of *P. noxius* in the transect area. Disease incidence at all sites was probably underreported, as *P. noxius* is usually transferred to healthy roots that touch diseased roots, stumps, or infected woody debris beneath the soil (Singh et al. 1980, Bolland 1984, Hodges & Tenorio 1984, Chang 1996). Early infections with no obvious above ground signs or symptoms would not be reported in our survey.

Disease incidence was lowest in montane and ridge top transects, increasing slightly in primary valley, slope, coastal and Tafuna Plain sites (Table 1). The number of infected trees in the secondary valley sites was more than twice the other sites. Webb and Fa'amu (1999) reported that catastrophic or repeated disturbances such as occur in secondary valleys affect stem density and species richness. These changes may in turn affect disease incidence (Bloomberg 1990). Other possible influences include edaphic factors, host susceptibility, aggregation of susceptible species, and virulence of the fungus.

Bloomberg (1990) cited high stocking levels

(stem density) and the increased probability of root contact in Douglas fir plantations on Vancouver Island, Canada, as a major cause of the spread of *Phellinus weirii* (Murill) R. L. Gilbertson. In tropical plantations, spread of *P. noxius* is usually within rows where spacing is closer, as opposed to between rows spaced farther apart (Neil 1986, Nandris et al. 1987). Webb et al. (1999) reported stem densities in DMWR plots were highest on ridges, possibly due to repeated thinning and shearing of trees caused by environmental disturbances, such as hurricanes and severe tropical storms. The opportunity for infection to spread by root-to-root contact should be greater on ridges where trees are closer together but in our surveys it was one of the least affected areas.

Stem densities were lower in primary valley sites but basal area measurements were higher than for either ridge or slope vegetation (Webb et al. 1999). In the same study on Douglas fir stem densities, Bloomberg (1990) also cites large average tree diameters as an important factor in *P. weirii* advance due to large root systems. In the DMWR plots, trees with large diameters (>50 cm) were the least frequent but had the highest proportion of brown root rot disease (Table 3).

Ridge soils are generally moist and well drained (Nakamura 1984), a condition favoring *P. noxius* infection (Bolland 1984, Chang 1996). The heavily infected secondary valley sites, Malaeimi I and II, are reported to be wet, poorly drained soils with a high water table (Nakamura 1984). According to Chang (1996), however, only flooded soils significantly limit *P. noxius* survival in infected wood: at soil matrix water potentials lower than -0.25 MPa, survival was 80-90 percent after two years. Most infected trees at Malaeimi I were either located along a dry, stony, well-drained streambed, or associated with surface roots (Figure 1b). We recorded no infections from the wettest portion of the Malaeimi II strip transect (Figure 1a).

Species richness is greater on ridge tops than in

other vegetation types of American Samoa (Webb et al. 1999). Nandris et al. (1987) suggest mixed stands inhibit root rot in tropical forests due to different above- and below ground structures, ecology and disease susceptibility. These conditions produce equilibrium between forest trees and fungi, with root rot maintained between 2-7 percent (Nandris et al. 1987): disease incidence in primary vegetation (DMWR plots) measured 3.2 percent. In spite of high stem densities and soils more favorable to *P. noxius*, a greater diversity of tree species on ridge tops may be responsible for the lower incidence and distribution of brown root rot disease.

The high number of infected trees in secondary valleys on Tutuila Island may be due to the relative abundance and aggregation of a few susceptible species, including *M. fatua*, *H. tiliaceus* and *D. samoense*. According to Webb et al. (1999), *M. fatua* was the most abundant species in the lowland forests of American Samoa. Neil (1986) found 40 percent of infected trees in Pentecost Island's natural forest were *M. fatua* and *M. fatua* var. *papuana*. He suggested this species was "preferentially colonized" by *P. noxius*. In mahogany plantations of Fiji, Singh and co-workers (1980) identified 15 native forest species as sources of infection in 44 disease centers: infection in 18 centers was initiated by *Myristica castaneaefolia* A. Gray. In our surveys, *M. fatua* accounted for 25 percent of all diseased trees and 23 percent of *P. noxius* infections in secondary valleys.

Single-species cropping systems are known for their susceptibility to disease (Fry 1982). Of 62 infection centers at all survey sites on Tutuila Island, 72 percent were either composed of a single tree species or a majority of the same species. Bloomberg (1990) reported a higher incidence of *P. weirii* in Canadian plantations where Douglas fir was dominant than in plantations with mixed stands. In the South Pacific, the incidence and spread of *P. noxius* is usually reported for and most severe in single-species plantations on cleared forest sites (Thrower 1965, Singh et al.

1980, Bolland 1984, Neil 1986, Ivory 1990). Webb and Fa'aumu (1999) suggest clearing of tropical forests by natural or human means leads to reestablishment by a few successional species. In secondary forests, *H. tiliaceus* is an important colonizing species (Webb et al. 1999) and *D. samoense* may dominate alluvial valleys (Whistler 1994). The high percentage of infection among these species and *M. fatua* could emphasize their prevalence in disturbed communities, susceptibility to *P. noxius*, virulence of the pathogen in these sites, or a combination of these factors.

Based on their findings, Nandris et al. (1987) believed infection of rubber trees in Africa was affected by differences in susceptibility within *Hevea* sp. and by intraspecific variability in pathogenicity of *P. noxius* isolates. In contrast to these findings, Chang (1995) tested 12 isolates of *P. noxius* on 9 host species and found no host specificity demonstrated by the fungus. Similar testing is needed to determine differences in host susceptibility to *P. noxius* in American Samoa, intraspecific variation in pathogenicity of fungal isolates, or both.

No large centers of brown root rot disease have been observed or reported on Tutuila Island. However, the relatively high levels of infection in disturbed forest sites and the broad host range of *P. noxius* suggest future problems for landscape plantings and local agriculture as people clear forest land to build houses and plant crops.

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REFERENCES

- Adaskaveg, J. E., and Ogawa, J. M. 1990. Wood decay pathology of fruit and nut trees in California. *Plant Disease* 74(5): 341-352.
- Ann, P. J., Lee, H. L., and Tsai, J. N. 1999. Survey of brown root disease of fruit and ornamental trees caused by *Phellinus noxius* in Taiwan. *Plant Pathology Bulletin* 8(2). (abstr.)
- Bloomberg, W. J. 1990. Effect of stand conditions on advance of *Phellinus weirii* in Douglas-fir plantations. *Phytopathology* 80: 553-559.
- Bolland, L. 1984. *Phellinus noxius*: cause of a significant root-rot in Queensland hoop pine plantations. *Australian Forestry* 47(1): 2-10.
- Chang, T. T. 1995. Decline of nine tree species associated with brown root rot caused by *Phellinus noxius* in Taiwan. *Plant Disease* 79: 962-965.
- Chang, T. T. 1996. Survival of *Phellinus noxius* in soil and in the roots of dead host plants. *Phytopathology* 86: 272-276.
- Chang, T. T. , and Yang, W. W. 1998. *Phellinus noxius* in Taiwan: distribution, host plants and the pH and texture of the rhizosphere soils of infected hosts. *Mycological Research* 102(9): 1085-1088.
- Cole, T. G., McKay, N., Whitsell, C. D., Ambacher, A. H., and Whistler, W. A. 1988. Vegetation survey and forest inventory, American Samoa. Pacific Southwest Forest and Range Experiment Station, Berkeley, CA.
- Corner, E. J. H. 1932. The identification of the brown-root fungus. *The Gardens' Bulletin, Straits Settlements* 5(12): 317-350.
- Fidalgo, O. 1968. *Phellinus pachyphloeus* and its allies. *Memoirs of the New York Botanical Garden* 17(2): 109-147.
- Fry, W. E. 1982. *Principles of Plant Disease Management*. Academic Press, Inc., New York.
- Greenwood, J. J. D. 1996. Basic techniques. In W. J. Sutherland (Ed.). *Ecological Census Techniques: A Handbook*, pp. 11-110. Cambridge University Press, Cambridge, UK.
- Haase, P. 1995. Spatial pattern analysis in ecology based on Ripley's K-function: Introduction and methods of edge correction. *Journal of Vegetation Science* 6: 575-582.
- Hodges, C. S., and Tenorio, J. A. 1984. Root disease of *Delonix regia* and associated tree species in the Mariana Islands caused by *Phellinus noxius*. *Plant Disease* 68: 334-336.

- Ivory, M. H. 1990. Brown root-rot of tropical forest trees in the South-West Pacific region. In C. Hutacharern, K. G. MacDicken, M. H. Ivory, and K. S. S. Nair (Eds.). Proceedings of the IUFRO Workshop on Pests and Diseases of Forest Plantations, pp. 100-106. Food and Agricultural Organization of the United Nations, Bangkok, Thailand.
- McKenzie, E. H. C. 1996. Fungi, bacteria and pathogenic algae on plants in American Samoa, Technical Paper No. 206. South Pacific Commission, Noumea, New Caledonia.
- Nakamura, S. 1984. Soil survey of American Samoa. USDA Soil Conservation Service, US Government Printing Office, Washington, DC.
- Nandris, D., Nicole, M., and Geiger, J. P. 1987. Root rot disease of rubber trees. *Plant Disease* 71(4): 298-306.
- Neil, P. E. 1986. A preliminary note on *Phellinus noxius* root rot of *Cordia alliodora* plantings in Vanuatu. *European Journal of Forest Pathology* 16: 274-280.
- Nicole, M., Chamberland, H., Rioux, D., Xixuan, X., Blanchette, R. A., Geiger, J. P., and Ouellette, G. B. 1995. Wood degradation by *Phellinus noxius*: ultrastructure and cytochemistry. *Canadian Journal of Microbiology* 41: 253-265.
- Pegler, D. N., and Waterston, J. M. 1968. *Phellinus noxius*, CMI Descriptions of Pathogenic Fungi and Bacteria, No. 195. Commonwealth Mycological Institute, Kew, Surrey, England.
- Ripley, B. D. 1981. *Spatial Statistics*. J. Wiley, New York.
- Singh, S., Bola, I., and Kumar, J. 1980. Diseases of plantation trees in Fiji Islands: I. Brown root rot of mahogany (*Swietenia macrophylla* King). *Indian Forester* 106: 526-532.
- Thrower, L. B. 1965. Parasitism of cacao by *Fomes noxius* in Papua³/₄New Guinea. *Tropical Agriculture, Trinidad* 42(1): 64-67.
- Trujillo, E. E. 1971. The breadfruit diseases of the Pacific Basin, Information Document No. 27. South Pacific Commission, Noumea, New Caledonia.
- Volk, R. D., Knudsen, P. A., Kluge, K. D., and Herdrich, D. J. 1992. Towards a territorial conservation strategy and the establishment of a conservation areas system for American Samoa. Le Vaomatua, Inc., Pago Pago, American Samoa.
- Webb, E. L., and Fa'aumu, S. 1999. Diversity and structure of tropical rain forest of Tutuila, American Samoa: effects of site age and substrate. *Plant Ecology* 144: 257-274.
- Webb, E. L., Brooks, J. S., and Jensen, M. L. 1999. Effects of topography on rainforest tree community structure and diversity in American Samoa, and implications for frugivore and nectarivore populations. *Journal of Biogeography* 26: 887-897.

Whistler, W. A. 1993. Botanical survey of the Ottoville lowland forest, Tafuna, Tutuila, American Samoa. Dept. of Marine and Wildlife Resources, Pago Pago, Am. Samoa. (unpublished)

Whistler, W.A. 1994. Botanical inventory of the proposed Tutuila and Ofu units of the National Park of American Samoa. Technical Report No. 87, Cooperative National Park Resources Studies Unit, University of Hawai'i at Manoa, Honolulu, HI.

Wingert, E.A. (Project Director). 1981. A coastal zone management atlas of American Samoa. University of Hawai'i Cartographic Laboratory, Honolulu, HI.

Table 1. Vegetation and soil types, and percent brown root rot disease (*Phellinus noxius*) for four established plots and 16 strip transects on Tutuila Island, American Samoa.

Site ^a	Vegetation Type ^b	Soil ^c	<i>P. noxius</i> ^d
1. Alava, DMWR	R, S, V	4	5.6
2. Amalau, DMWR	S, V	4, 2	8.6
3. Maloata, DMWR	R, S	4	7.1
4. Vatia, DMWR	R, S, V	4	7.1
5. Matafao Peak	M	4	0.6
6. Maatula	M	4	1.2
7. Olo	R	2	1.5
8. Tuasina	R	4	1.2
9. Faga'alu	S	4	2.2
10. A'asutuai	S	4	2.8
11. Malaeloa	V	8	2.5
12. Nu'uuli	V	2	3.1
13. Vatia I	L	2	4.3
14. Taputapu	L	4	2.5
15. Tafuna	P	32	7.4
16. Vatia II	2° R	4, 2	1.2
17. Maloata	2° V	4	15.1
18. Malaeimi I	2° V	8	18.8
19. Malaeimi II	2° V	8	7.1
20. Pago Airport	2° P	33	0

^a DMWR = American Samoa Government Department of Marine and Wildlife Resources 1.2 ha plots. Other sites listed are 0.25 ha strip transects.

^b Vegetation types (Whistler 1994): R = ridge, S = slope, V = valley bottom, M = montane (>350 m elevation), L = littoral, P = Tafuna Plain, 2° = secondary or disturbed sites.

^c Soil type and slope (see text for description): 4 = Lithic-Hapludolls-Rock outcrop association, very steep; 2 = Aua very stony silty clay loam, 30-60%; 32 = Tafuna extremely stony muck, 3-15%; 8 = Leafu silty clay, 0-3%; 33 = Troporthents, 0-6% (Nakamura 1984).

^d Percent of the total number of trees infected with *Phellinus noxius* from all sites (324). Note: area of DMWR plots is five times greater than strip transects.

Table 2. Tree species infected by *Phellinus noxius* on Tutuila Island, American Samoa, January to May 2001. Included are host family, authority, total number of infected trees for each species and minimum, maximum and average diameter at breast height.

Host, Authority ^a	No.	dbh (cm) ^b
Anacardiaceae		
<i>Rhus taitensis</i> Guillemain	11	(9-86) 45
<i>Spondias dulcis</i> L.	4	(26-79) 63
Annonaceae		
<i>Cananga odorata</i> (Lam.) Hook & Thoms.	8	(13-44) 29
Apocynaceae		
<i>Cerbera manghas</i> L.	2	(8.3-38.5) 23
Barringtoniaceae		
<i>Barringtonia asiatica</i> (L.) Kurz	3	(4.5-64.5) 25
<i>Barringtonia samoensis</i> A. Gray	8	(11.5-72) 25
Boraginaceae		
<i>Cordia aspera</i> Forst. f.	1	13.5
Burseraceae		
<i>Canarium harveyi</i> Seem.	5	(6-41.5) 18
Clusiaceae		
<i>Calophyllum neo-ebudicum</i> Guillaumin	6	(58-190) 132
Combretaceae		
<i>Terminalia richii</i> A. Gray	1	17
Ebenaceae		
<i>Diospyros samoensis</i> A. Gray	1	5
Euphorbiaceae		
<i>Flueggea flexuosa</i> Marg.	3	(12-15) 13.5
<i>Glochidion ramiflorum</i> Forst.	1	11
<i>Macaranga harveyana</i> (Muell. Arg.) Muell. Arg.	20	(2-34) 15
<i>Macaranga stipulosa</i> Muell. Arg.	10	(14-38) 24
Fabaceae		
<i>Adenantha pavonina</i> L.	1	15
<i>Inocarpus fagifer</i> (Parkinson) Fosb.	9	(5-89) 31
<i>Intsia bijuga</i> (Colebr.) Kuntze	7	(2.5-57) 20
<i>Samanea saman</i> (Jacq.) Merr.	1	93
Hernandiaceae		
<i>Hernandia nymphaeifolia</i> (Presl) Kub.	1	38
Malvaceae		

<i>Hibiscus tiliaceus</i> L.	29	(6-68) 21
Meliaceae		
<i>Dysoxylum samoense</i> A. Gray	45	(2-160) 25
Moraceae		
<i>Ficus obliqua</i> Forst. f.	3	(89-123) 109
<i>Ficus tinctoria</i> Forst. f.	1	5
<i>Ficus</i> sp.	1	5
Myristicaceae		
<i>Myristica fatua</i> Houtt.	69	(3-140) 19
Myrtaceae		
<i>Syzygium inophylloides</i> (A. Gray) C. Muell.	2	(15-51) 33
<i>Syzygium</i> sp.	1	19
Rhizophoraceae		
<i>Crossostylis biflora</i> Forst.	1	17
Rubiaceae		
<i>Morinda citrifolia</i> L.	3	(2.5-10) 5
<i>Neonauclea forsteri</i> (Seem. ex Havil.) Merr.	1	102
Sapotaceae		
<i>Planchonella grayana</i> St. John	2	(57-83) 66
<i>Planchonella samoensis</i> H.J. Lam ex Christoph.	20	(5-110) 42
Sapindaceae		
<i>Elattostachys falcata</i> (A. Gray) Radlk.	2	(33.4-35) 34
<i>Pometia pinnata</i> Forst.	3	(5-22) 12
Urticaceae		
<i>Pipturus argenteus</i> (Forst. f.) Wedd.	2	(11-22) 17
Unidentified	25	(4-80) 24

^a Nomenclature after Whistler (1994) and Farr et al. (1995).

^b Minimum, maximum and average diameter at breast height.

Table 3. Percentage of trees in three diameter classes at American Samoa Government Department of Marine and Wildlife Resources sites Alava, Amalau and Vatia (total) followed by the percent of infected trees in each class surveyed at these sites (infected).

Diameter Class ^a	10-30 cm	30-50 cm	>50 cm
Total	81%	13%	6%
Infected	55%	21%	23%

^a Webb and Fa'aumu (1999).

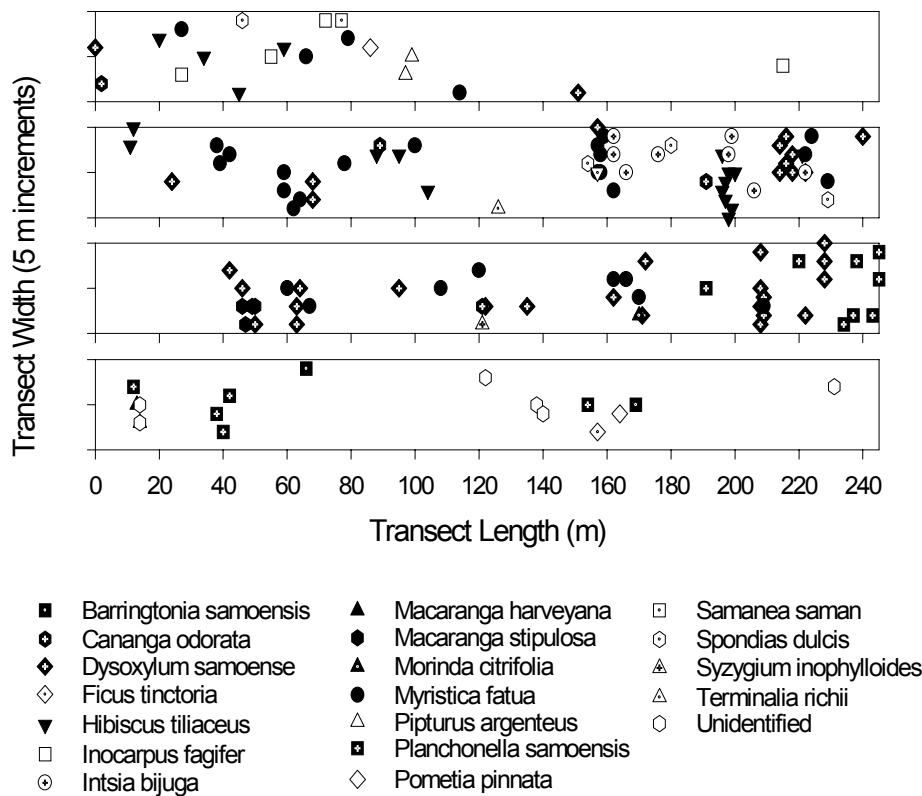


FIGURE 1. Strip transects at secondary (disturbed) lowland forest sites, Tutuila Island, American Samoa. Each transect is 10 m x 245 m (0.25 ha); symbols represent tree species and their approximate location within transects. Sites include (1a) Malaeimi Valley II, (1b) Malaeimi Valley I, (1c) Maloata Valley, and (1d) Tafuna Plain.

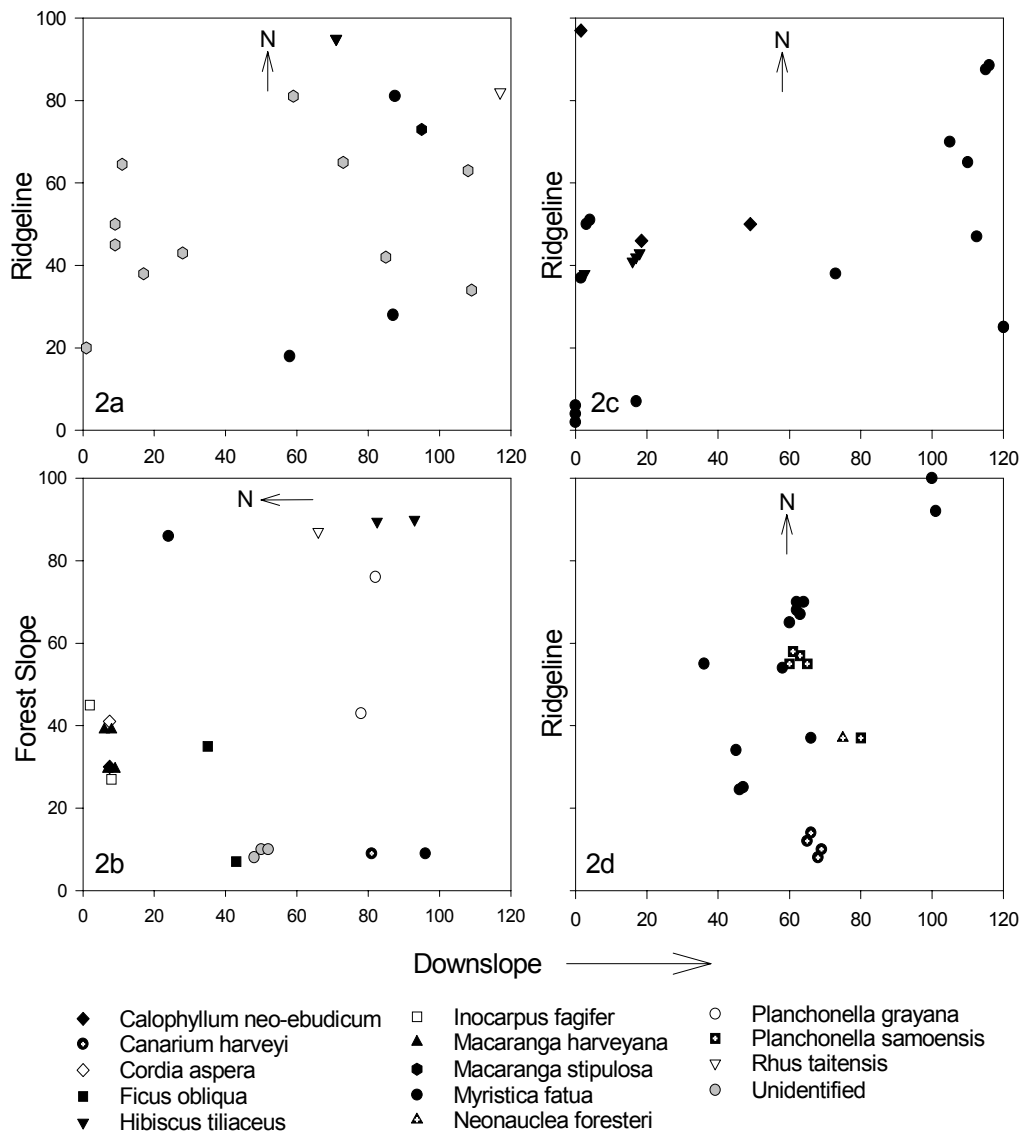


FIGURE 2. Four permanent forest plots on Tutuila Island managed by the American Samoa Government Department of Marine and Wildlife Resources (DMWR). Plots are relatively free of human disturbance and measure 100 m x 120 m (1.2 ha). They include ridgelines, slopes and valleys with the exception of Amalau, sited below the ridgeline and Maloata, which does not include a valley bottom. DMWR plots are located at (2a) Alava, (2b) Amalau, (2c) Maloata, and (2d) Vatia.